



New access to the synthetic building block L-aspartic acid β -semialdehyde via Grignard reaction

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Received 14 November 2000; accepted 5 December 2000

Abstract

A new, efficient, and inexpensive synthesis of protected L-aspartic acid β -semialdehyde has been developed starting from L-glutamic acid via a substituted L-allylglycine derivative as intermediate. The key step of the reaction sequence was a strongly solvent-dependent Grignard reaction of an L-glutamic acid semiesther. The desired regioselective addition to the C-5 ester group was achieved in 1,2-dimethoxyethane while reactions in diethyl ether gave products resulting from additional attack at the carboxylic acid functionality. © 2001 Elsevier Science Ltd. All rights reserved.

1. Introduction

L-Aspartic acid β -semialdehyde (**1**) is a natural non-proteinogenic amino acid and a key intermediate in the biosynthesis of the essential amino acids L-lysine, L-methionine, L-threonine, and L-isoleucine in higher plants and bacteria.¹ It also represents a relevant synthetic intermediate. The potential access to a variety of polyfunctional non-proteinogenic and unnatural amino acids using **1** has been demonstrated.² Further applications include the total syntheses of rhizobitoxin³ and sinefungin,⁴ and also of the phytotoxic agent D,L- γ -hydroxyphosphinothricine via protected, racemic **1**.⁵

An established route to **1** employs ozonolysis of L-allylglycine⁶ or its derivatives,⁷ which in turn is accessible by malonester syntheses⁸ and via glycine-derived⁹ or other stable imines¹⁰ followed by enzyme-catalyzed racemate resolution. Other known methods start from the chiral pool and proceed through oxidation of L-homoserines, which can be prepared from L-methionine,² and by controlled reduction¹¹ or, preferably, by reduction–oxidation¹² of L-aspartic acid. However, *N*-alkoxycarbonyl derivatives of homoserines reportedly have the tendency of lactonization.¹³ Furthermore, many reducing agents are not compatible with widely-used protecting

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groups.¹⁴ Common oxidation methods for the synthesis of some aldehydes have been successfully utilized,¹⁵ although they led to unsatisfactory results for other authors.¹⁶ None of these methods is predominant in the literature.

The need for enantiomerically pure **1** for biochemical and organic-preparative studies demands an efficient, inexpensive synthetic route of general validity. We describe a method that combines the strengths of both known approaches using the chiral pool of the biogenic amino acids and the mild ozonolysis of an L-allylglycine analogue by an overall Barbier–Wieland degradation. To avoid stability problems and because of its intended further use in natural product synthesis, the doubly protected derivative of **1**, *N*-ethoxycarbonyl-L-aspartic acid α -methyl ester **2**, had been chosen as a target molecule (see Fig. 1).

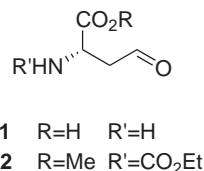
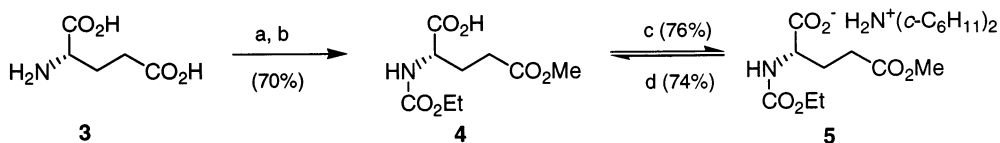


Figure 1.

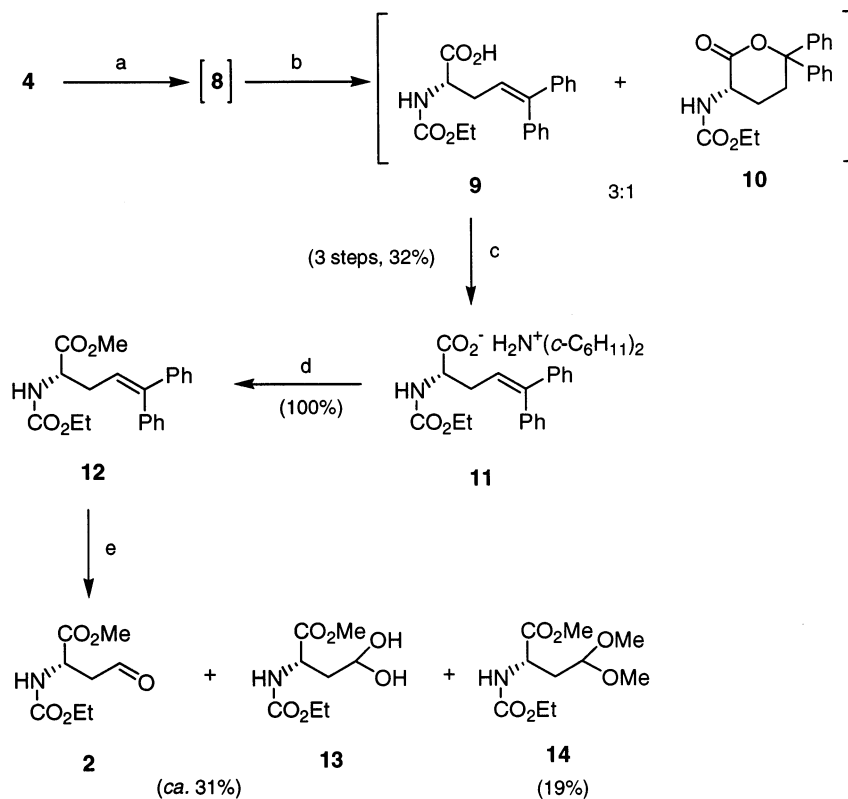
2. Results and discussion

L-Glutamic acid **3** served as starting material. The two acid functions were differentiated by selective esterification of the γ -carboxyl group to secure chemoselectivity in later stages in the synthesis. Esterification and subsequent protection of the amino functionality were carried out analogous to our established method for L-aspartic acid.¹⁷ The NMR-spectroscopically pure, yet oily semiester **4** was obtained in high yield. It was further characterized as its dicyclohexylammonium salt **5**, which served as a storage compound because of its stability (see Scheme 1).



Scheme 1. (a) MeOH, HCl, 20 h, rt; then NaHCO₃; (b) NaHCO₃, ClCO₂Et, 16 h, rt; (c) NH(*c*-C₆H₁₁)₂, Et₂O; (d) NaHCO₃, Et₂O; then H₂SO₄, Et₂O

Grignard reagents are known to add to ester groups but usually not to free acid or carbamate functionalities, even though we recently observed side products arising from attack at the acid functionality of *N*-alkoxycarbonyl-L-aspartic acid β -methyl ester.¹⁸ Reaction with semiester **4** was expected to mainly afford a tertiary alcohol that would dehydrate to a substituted L-allylglycine derivative under mild conditions. Phenylmagnesium bromide was chosen as the Grignard reagent to favor subsequent acid-catalyzed dehydration because of the stabilizing effect of the phenyl groups. Four equivalents of Grignard reagent are needed for the reaction: two equivalents to abstract the acidic protons of the carbamate nitrogen and carboxylic acid functions, and the other two for the nucleophilic addition onto the ester functionality.



Scheme 2. (a) PhMgBr, DME, 2 h, reflux; then H₂O; (b) H₂SO₄, pH 1.0, 0°C; (c) NH(*c*-C₆H₁₁)₂, Et₂O; (d) Me₂SO₄, MeOH, 2 h, steam bath; (e) O₃, CH₂Cl₂/MeOH, -60°C; then Me₂S, 14 h, rt

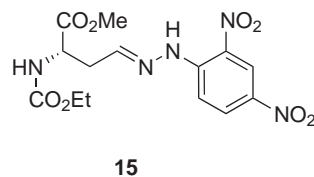


Figure 3.

costly reagents or possibly harsh oxidizing/reducing conditions, and consists of only a few steps. It therefore represents a superior alternative to the established methods in the literature. However, the relatively low pH that is necessary for the dehydration excludes acid-sensitive *N*-protecting groups. The key to the success of the synthesis was the control of the regioselectivity in the reaction of the *N*-protected semiester **4** with phenylmagnesium bromide, where using DME as the solvent allowed the nucleophilic addition to be directed successfully towards the ester group. Exploiting this selective reactivity may enhance the applicability of Grignard reactions in natural product synthesis.

4. Experimental

4.1. General methods

Organic solvents were purified before use. Diethyl ether (Et₂O) and 1,2-dimethoxyethane (DME) were freed of peroxides by storage over KOH. After subsequent drying over sodium until positive benzophenone reaction, both ethers were distilled under an inert argon atmosphere. Preparations of phenylmagnesium bromide were also carried out under argon. TLC analysis was performed using Polygram Sil G/UV₂₅₄ plates from Macherey–Nagel. The *R_f* values of salts given are equivalent to those of the corresponding acids since acidic mobile phases have been used. Silica gel S (0.032–0.063 mm, 230–400 mesh) from Riedel–de Haën was used for flash chromatography. Melting points were determined on a Büchi 510 apparatus and are uncorrected. Optical rotations were measured with the automatic polarimeter AA 1000 from Technolab. IR spectra were recorded on a Perkin–Elmer FTIR 1750 instrument. ¹H and ¹³C NMR experiments were carried out on a Bruker AC 200 spectrometer operating at 200 and 50 MHz, respectively, using tetramethylsilane as the internal standard. Mass spectra were obtained on an HP5988 instrument from Hewlett–Packard, operated in the EI mode at 70 eV. Combustion analyses were performed by Beller Mikroanalytisches Labor, Göttingen (Germany).

4.2. *N*-Ethoxycarbonyl-*L*-glutamic acid γ -methyl ester **4**

L-Glutamic acid **3** (22.1 g, 150 mmol) was suspended in MeOH (400 mL). Addition of 37% aqueous HCl (45 mL, 540 mmol) resulted in a colorless solution that was stirred at 25°C for 20 h. H₂O (120 mL) and NaHCO₃ (46.0 g, 548 mmol) were added, and MeOH was evaporated in vacuo. H₂O was added to give a final volume of 250 mL followed by further NaHCO₃ (33.6 g, 400 mmol). Ethyl chloroformate (32.6 g, 300 mmol) was added dropwise to the solution that was then stirred for 16 h at 25°C. The solution was washed with Et₂O (3×50 mL) and the organic phases were discarded. The aqueous solution was cooled using an ice bath and acidified to pH 2 with cold 50% H₂SO₄. The product was extracted with Et₂O (4×150 mL). The combined ethereal phases were washed with H₂O (50 mL), dried over MgSO₄, and the solvent was evaporated to give semiester **4** as a colorless to slightly yellowish oil (24.6 g, 70%): *R_f* 0.23 [EtOAc/*n*-hexane/AcOH (30:60:1)]; ¹H NMR (CDCl₃) δ 1.18 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 1.86–2.08 (m, 1H, CHH'CHN), 2.08–2.30 (m, 1H, CHH'CHN), 2.43 (td, *J* = 7.7, 2.8 Hz, 2H, CH₂CO₂Me), 4.07 (q, *J* = 7.1 Hz, 2H, OCH₂), 4.20–4.42 (m, 1H, CH), 5.61 (br d, *J* = 8.1 Hz, 0.75H, NH), 6.54 (br d, *J* = 7.0 Hz, 0.25H, NH), 10.12 (br s, 1H, CO₂H); ¹³C NMR (CDCl₃) δ 14.3 (q, OCH₂CH₃), 27.2 (t, CH₂CHN), 30.0 (t, CH₂CO₂Me), 51.8 (q, OCH₃), 52.9 (d, CH), 61.4 (t, OCH₂), 156.5 (s, NHCO₂Et), 173.5 (s, CO₂Me), 175.3 (s, CO₂H).

4.3. Dicyclohexylammonium 5-methyl-(*S*)-*N*-ethoxycarbonylglutamate **5**

Compound **5** was prepared for characterization and storage purposes. Dicyclohexylamine (5.80 g, 32.0 mmol) was added to a solution of semiester **4** (7.46 g, 32.0 mmol) in Et₂O (100 mL). Ammonium salt **5** crystallized overnight in the refrigerator. The colorless crystals were filtered, washed with cold Et₂O and dried in vacuo (10.1 g, 76%): *R_f* 0.23 [EtOAc/*n*-hexane/AcOH (30:60:1)]; mp 127–128°C; [α]_D²⁰ +11.2 (*c* 1, MeOH); IR (KBr) ν_{\max} 3227, 3000–2400, 1737, 1719, 1631, 1547, 1448, 1406, 1374, 1266, 1169, 1075 cm⁻¹; ¹H NMR (CDCl₃) δ 1.03–2.52 (m, 24H, 10

ring-CH₂, CH₂CH₂CO₂Me), 1.17 (t, $J=7.1$ Hz, 3H, OCH₂CH₃), 2.81–3.02 (m, 2H, 2 ring-CH), 3.58 (s, 3H, OCH₃), 3.91–4.13 (m, 3H, OCH₂, NCHCO₂⁻), 5.60 (br d, $J=6.1$ Hz, 1H, NH), 8.20–9.70 (br, 2H, N⁺H₂); ¹³C NMR (CDCl₃) δ 14.6 (q, OCH₂CH₃), 4 \times 24.7 (t, ring-CH₂), 2 \times 25.0 (t, ring-CH₂), 28.9 (t, CH₂CHN), 4 \times 29.0 (t, ring-CH₂), 30.3 (t, CH₂CO₂Me), 51.4 (q, OCH₃), 2 \times 52.5 (d, ring-CH), 55.2 (d, NCHCO₂⁻), 60.4 (t, OCH₂), 156.1 (s, NHCO₂Et), 174.0 (s, CO₂Me), 175.4 (s, CO₂⁻). Anal. calcd for C₂₁H₃₈N₂O₆: C 60.85; H 9.24. Found: C, 60.81; H, 9.21.

To liberate semiester **4**, the ammonium salt **5** (7.50 g, 18.1 mmol) was mixed in a separatory funnel with an aqueous solution of NaHCO₃ (6.50 g, 77.4 mmol) in H₂O (70 mL) and Et₂O (35 mL). NaOH was added until the aqueous phase reached pH 9.5. The ethereal phase was separated, and the aqueous phase extracted with Et₂O (2 \times 35 mL) to remove dicyclohexylamine. The aqueous phase was acidified to pH 2 with 50% H₂SO₄ and extracted with Et₂O (4 \times 60 mL). The organic phases were dried over MgSO₄ and the solvent was evaporated to give semiester **4** (3.13 g, 74%).

4.4. Dicyclohexylammonium (S)-2-(ethoxycarbonylamino)-5,5-diphenyl-4-pentenoate **11**

Grignard reactions were carried out in Et₂O and DME at various stoichiometric ratios and using different reaction work-ups. Reactions in DME using a molar ratio of bromobenzene/semiester **4** of 10:1 gave the best results. The following procedure proved to be the most efficient.

The Grignard reagent was prepared by dropwise addition of a solution of bromobenzene (35.3 g, 225 mmol) in DME (40 mL) to a slurry of magnesium (5.83 g, 240 mmol) in DME (60 mL). The reaction started after heating the mixture. To complete the reaction, the mixture was heated under reflux for an additional 2 h. The Grignard reagent was transferred into a dropping funnel under an argon atmosphere, while the remaining precipitate and excess magnesium were removed by this procedure. The Grignard reagent was then added to a solution of semiester **4** (5.00 g, 21.4 mmol) in DME (75 mL) under vigorous stirring and sonication. The white suspension was heated under reflux for 2 h and it finally turned yellow. The mixture was hydrolyzed with H₂O (100 mL), acidified until pH 1.0 with H₂SO₄ (50%) under cooling of the flask with ice. The precipitate dissolved, and the red organic phase was separated. The aqueous, mixed phase was extracted with Et₂O (4 \times 100 mL). The extracts were combined with the previously separated organic phase and washed with aqueous NaHCO₃ solution of pH 8.0 (2 \times 25 mL) and then further washed with H₂O (25 mL). The ethereal phase was concentrated to a final volume of 30 mL and dicyclohexylamine (3.88 g, 21.4 mmol) was added. The mixture was allowed to crystallize overnight in the refrigerator, the precipitated product was then collected by filtration and recrystallized from CHCl₃/*n*-hexane to afford pure **11** (3.54 g, 32% from **4**): R_f 0.47 [EtOAc/*n*-hexane/AcOH (30:60:1)]; mp 140–141°C; $[\alpha]_D^{20} +36.5$ (c 1, MeOH); IR (KBr) ν_{\max} 3422, 3054, 3025, 3000–2400, 1708, 1631, 1595, 1557, 1539, 1495, 1444, 1420, 1397, 1242, 1062, 758, 701 cm⁻¹; ¹H NMR (CDCl₃) δ 1.00–2.03 (m, 23H, 10 ring-CH₂, CH₃), 2.48–3.06 [m, 4H, CH₂CHN, N⁺(CH₂)₂], 3.97–4.28 (m, 3H, OCH₂, NCHCO₂⁻), 5.62 (br d, $J=7.1$ Hz, 1H, NH), 6.21 [t, $J=7.0$ Hz, 1H, CHC(Ph)₂], 7.15–7.43 (m, 10H, CH_{ar}), 7.90–8.90 (br, 2H, N⁺H₂); ¹³C NMR (CDCl₃) δ 14.7 (q, CH₃), 4 \times 24.7 (t, ring-CH₂), 2 \times 25.0 (t, ring-CH₂), 4 \times 29.1 (t, ring-CH₂), 34.0 (t, CH₂CHN), 52.4 [d, N⁺(CH₂)₂], 55.6 (d, NCHCO₂⁻), 60.3 (t, OCH₂), 126.0 [d, CHC(Ph)₂], 126.7, 127.2, 127.9, 128.0, 129.9 (d, 10 \times CH_{ar}), 140.0, 142.8, 143.1 [s, 2 \times C_{ar}^{ipso}, C(Ph)₂], 155.9 (s, NHCO₂Et), 175.6 (s, CO₂⁻). Anal. calcd for C₃₂H₄₄N₂O₄: C, 73.81; H, 8.52. Found: C, 73.87; H, 8.42.

4.5. Methyl (S)-2-(ethoxycarbonylamino)-5,5-diphenyl-4-pentenoate **12**

A solution of ammonium salt **11** (2.09 g, 4.02 mmol) and dimethyl sulfate (506 mg, 4.02 mmol) in MeOH (20 mL) was heated in a steam bath for 2 h. H₂O (50 mL) was added and the solution extracted with Et₂O (4×50 mL). The combined ethereal phases were washed with saturated aqueous NaHCO₃ (50 mL) and then with H₂O (50 mL). The solvent was evaporated to give **12** as an oily, yellowish residue (1.42 g, 100%): *R*_f 0.48 [Et₂O/*n*-pentane (1:1)]; IR (film) ν_{\max} 3336, 3056, 3025, 2981, 2953, 1745, 1723, 1599, 1525, 1497, 1444, 1372, 1213, 1178, 1059, 762, 702 cm⁻¹; ¹H NMR (CDCl₃) δ 1.14 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 2.37–2.72 (m, 2H, CH₂CHN), 3.61 (s, 3H, OCH₃), 4.02 (q, *J* = 7.1 Hz, 2H, OCH₂), 4.32–4.47 (m, 1H, CHN), 5.16 (br d, *J* = 8.4 Hz, 1H, NH), 5.91 [t, *J* = 7.4 Hz, 1H, CHC(Ph)₂], 6.95–7.39 (m, 10H, CH_{ar}); ¹³C NMR (CDCl₃) δ 14.5 (q, OCH₂CH₃), 32.7 (t, CH₂CHN), 52.3 (q, OCH₃), 53.5 (d, CHN), 61.1 (t, OCH₂), 122.6 [d, CHC(Ph)₂], 127.2, 127.3, 127.7, 128.1, 128.3, 129.6 (d, 10×CH_{ar}), 139.3, 142.0, 145.4 [2×C_{ar}^{ipso}, C(Ph)₂], 155.9 (s, NHCO₂Et), 172.3 (s, CO₂Me); MS (EI) *m/z* (%) 353 (2) [M⁺], 294 (52) [M⁺–CO₂CH₃], 205 (88), 178 (17), 165 (30), 144 (16), 115 (23), 90 (84), 77 (9), 59 (6) [CO₂CH₃⁺], 29 (56), 28 (100) [C₂H₄^{•+}].

4.6. Ozonolysis of methyl (S)-2-(ethoxycarbonylamino)-5,5-diphenyl-4-pentenoate **12**

Ester **12** (1.41 g, 3.99 mmol) was dissolved in a mixture of CH₂Cl₂/MeOH (2:1) (300 mL) and ozonized at –60°C. Dimethyl sulfide (1 mL) was added and the solution stirred at room temperature overnight. The solvent was removed in vacuo. Flash chromatography of the residue gave semialdehyde **2**, partially hydrated to **13**, and its dimethyl acetal **14**.

4.7. Mixture of N-ethoxycarbonyl-L-aspartic acid α -methyl ester β -semialdehyde **2** and N-ethoxycarbonyl-L-aspartic acid α -methyl ester β -semialdehyde hydrate **13**[†]

Slightly yellowish oil (250 mg, ca. 31%); *R*_f 0.27 [Et₂O/*n*-pentane (2:1)]; IR (film) ν_{\max} 3362, 2982, 2940, 2720, 1740, 1723, 1700, 1531, 1440, 1374, 1221, 1180, 1066 cm⁻¹; ¹H NMR (CDCl₃) for **2**: δ 1.17 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 3.02 (t, *J* = 5.0 Hz, 2H, CH₂CHN), 3.68 (s, 3H, OCH₃), 4.05 (q, *J* = 7.1 Hz, 2H, OCH₂), 4.58 (dt, *J* = 8.4, 5.0 Hz, 1H, CHN), 5.76 (br d, *J* = 8.4 Hz, 1H, NH), 9.66 (s, 1H, CHO); ¹H NMR (CDCl₃) for **13**: δ 1.17 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 1.98–2.10 (m, 2H, CH₂CHN), 3.68 (s, 3H, OCH₃), 4.05 (q, *J* = 7.1 Hz, 2H, OCH₂), 4.37–4.48 (m, 1H, CHN), 5.20–5.32 [m, 1H, CH(OH)₂], 5.65 (br d, *J* = 7.9 Hz, 1H, NH); ¹³C NMR (CDCl₃) for **2**: δ 14.4 (q, OCH₂CH₃), 45.8 (t, CH₂CHN), 48.8 (d, CHN), 52.8 (q, OCH₃), 61.4 (t, OCH₂), 156.1 (s, NHCO₂Et), 171.2 (s, CO₂Me), 199.3 (d, CHO); ¹³C NMR (CDCl₃) for **13**: δ 14.4 (q, OCH₂CH₃), 35.1 (t, CH₂CHN), 50.5 (d, CHN), 52.5 (q, OCH₃), 61.4 (t, OCH₂), 98.5 [d, CH(OH)₂], 156.1 (s, NHCO₂Et), 172.5 (s, CO₂Me); MS (EI) *m/z* (%) 203 (0.1) [M⁺ for **2**], 175 (2), 171 (8), 144 (100) [(M⁺–CO₂CH₃) for **2**], 130 (10), 115 (11), 102 (6), 88 (6), 72 (17), 59 (6), 44 (52) [CH₂=CH–OH^{•+}], 29 (25).

[†] NMR signal assignments were made with aid of 2D NMR correlation experiments and published NMR data for similar compounds.

4.8. Methyl (S)-2-(ethoxycarbonylamino)-4,4-dimethoxybutanoate **14**

Slightly yellowish oil (187 mg, 19%); R_f 0.49 [Et₂O/*n*-pentane (2:1)]; IR (film) ν_{\max} 3400, 2980, 2954, 2820, 1745, 1713, 1542, 1440, 1373, 1224, 1185, 1130–1000 cm⁻¹; ¹H NMR (CDCl₃) δ 1.17 (t, $J=7.1$ Hz, 3H, OCH₂CH₃), 1.90–2.18 (m, 2H, CH₂CHN), 3.24 (s, 3H, CH₃OCHOCH₃'), 3.27 (s, 3H, CH₃OCHOCH₃'), 3.67 (s, 3H, CO₂CH₃), 4.05 (q, $J=7.1$ Hz, 2H, OCH₂), 4.28–4.46 (m, 2H, CHCH₂CH), 5.57 (br d, $J=7.9$ Hz, 1H, NH); ¹³C NMR (CDCl₃) δ 14.4 (q, OCH₂CH₃), 34.8 (t, CH₂CHN), 50.7 (d, CHN), 52.2, 53.2, 53.6 [q, CO₂CH₃, CH(OCH₃)₂], 61.0 (t, OCH₂), 102.0 [d, CH(OMe)₂], 156.3 (s, NHCO₂Et), 172.8 (s, CO₂Me); MS (EI) m/z (%) 249 (0.01) [M⁺], 218 (2), 190 (5) [M⁺-CO₂CH₃], 161 (4), 158 (9), 129 (33), 88 (6), 75 (100) [CH(OCH₃)₂]⁺, 59 (5), 29 (6).

4.9. N-Ethoxycarbonyl-L-aspartic acid α -methyl ester β -semialdehyde-(2,4-dinitro)phenylhydrazone **15**

A freshly prepared solution of the mixture of **2** and **13** (100 mg, ca. 479 μ mol) in EtOH (1 mL) was added to a solution of 2,4-dinitrophenylhydrazine (400 mg, 2.02 mmol) in concentrated H₂SO₄ (2 mL) and H₂O (25 mL). The resultant precipitate was filtered, washed with H₂O, recrystallized from EtOAc/*n*-hexane, and dried in vacuo. Yellow crystals of **15** were obtained (20.5 mg, ca. 11%); mp 167–168°C; $[\alpha]_D^{20}$ +40.2 (c 0.5, CHCl₃); ¹H NMR (CDCl₃) δ 1.18 (t, $J=7.1$ Hz, 3H, OCH₂CH₃), 2.80–3.06 (m, 2H, CH₂CHN), 3.73 (s, 3H, OCH₃), 4.07 (q, $J=7.1$ Hz, 2H, OCH₂), 4.56–4.73 (m, 1H, NCHCO₂Me), 5.41 (br d, $J=8.1$ Hz, 1H, NHCO₂Et), 7.44 (t, $J=4.4$ Hz, 1H, CHNNH), 7.77 (d, $J=9.6$ Hz, 1H, CH_{ar}⁽⁶⁾), 8.28 (dd, $J=9.6, 2.6$ Hz, 1H, CH_{ar}⁽⁵⁾), 9.05 (d, $J=2.6$ Hz, 1H, CH_{ar}⁽³⁾), 11.0 (s, 1H, NNH); ¹³C NMR (CDCl₃) δ 14.5 (q, OCH₂CH₃), 35.4 (t, CH₂CHN), 51.2 (d, NCHCO₂Me), 52.9 (q, OCH₃), 61.5 (t, OCH₂), 116.4 (d, CH_{ar}), 123.4 (d, CH_{ar}), 129.3 (s, C_{ar}^{quart.}), 130.2 (d, CH_{ar}), 138.4 (s, C_{ar}^{quart.}), 144.9 (s, C_{ar}^{quart.}), 146.5 (d, CHNNH), 156.0 (s, NHCO₂Et), 171.7 (s, CO₂Me). Anal. calcd for C₁₄H₁₇N₅O₈: C, 43.87; H, 4.47. Found: C, 44.65; H, 4.77.

Acknowledgements

The authors thank the Deutsche Forschungsgemeinschaft and the Fond der Chemischen Industrie for financial support.

References

1. Hunt, S. In *Chemistry and Biochemistry of the Amino Acids*; Barrett, G. C., Ed. The non-protein amino acids. Chapman and Hall: London, 1985; pp. 55–138.
2. Baldwin, J. E.; Flinn, A. *Tetrahedron Lett.* **1987**, 28, 3605–3608.
3. Keith, D. D.; Tortora, J. A.; Ineichen, K.; Leimgruber, W. *Tetrahedron* **1975**, 31, 2633–2636.
4. Mock, G. A.; Moffatt, J. G. *Nucleic Acids Res.* **1982**, 10, 6223–6234.
5. Walker, D. M.; McDonald, J. F.; Franz, J. E.; Logusch, E. W. *J. Chem. Soc., Perkin Trans. 1* **1990**, 659–666.
6. Black, S.; Wright, N. G. *J. Biol. Chem.* **1955**, 213, 39–50.
7. Tudor, D. W.; Lewis, T.; Robins, D. J. *Synthesis* **1993**, 1061–1062.
8. Albertson, N. F. *J. Am. Chem. Soc.* **1946**, 68, 450–453.

9. Stork, G.; Leong, A. Y. W.; Touzin, A. M. *J. Org. Chem.* **1976**, *41*, 3491–3493.
10. Baldwin, J. E.; Bradley, M.; Turner, N. J.; Adlington, R. M.; Pitt, A. R.; Sheridan, H. *Tetrahedron* **1991**, *47*, 8203–8222.
11. Ito, A.; Takahashi, R.; Baba, Y. *Chem. Pharm. Bull.* **1975**, *23*, 3081–3087.
12. Stanfield, C. F.; Parker, J. E.; Kanellis, P. *J. Org. Chem.* **1981**, *46*, 4797–4798.
13. Barlos, K.; Theodoropoulos, D. *Z. Naturforsch., Teil B* **1982**, *37*, 886–888.
14. Stanfield, C. F.; Parker, J. E.; Kanellis, P. *J. Org. Chem.* **1981**, *46*, 4799–4800.
15. Faust, J.; Schreiber, K.; Ripperger, H. *Z. Chem.* **1984**, *24*, 330–331.
16. Walker, D. M.; McDonald, J. F.; Logusch, E. W. *J. Chem. Soc., Chem. Commun.* **1987**, 1710–1711.
17. Uzar, H. C. *Synthesis* **1991**, 526–528.
18. Brinkmann, T.; Gilg, A.; Hamm, A.; Lüscher, H.; Morbach, G.; Uzar, H. C. *Tetrahedron: Asymmetry* **2000**, *11*, 3827–3836.
19. Nützel, K. In *Methoden der Organischen Chemie (Houben-Weyl)*, 4th ed.; Müller, E., Ed.; Organo-magnesium-Verbindungen. Georg Thieme: Stuttgart, 1973; Vol. XIII/2a, pp. 47–527.
20. Lüscher, H. Diplomarbeit, Universität-Gesamthochschule Siegen, 1997.
21. Cohen, H. L.; Wright, G. F. *J. Org. Chem.* **1953**, *18*, 432–446.
22. Ohki, S.; Hamaguchi, F. *Yakugaku Zasshi* **1965**, *85*, 971–975.